

EpiQuik™ Global Acetyl Histone H4-K16 Quantification Kit (Fluorometric)

Base Catalog # P-4027

PLEASE READ THIS ENTIRE USER GUIDE BEFORE USE

The EpiQuik™ Global Acetyl Histone H4-K16 Quantification Kit (Fluorometric) is suitable for specifically measuring global histone H4-K16 acetylation using a variety of mammalian cells (human, mouse, etc.) including fresh and frozen tissues, and cultured adherent and suspension cells.



KIT CONTENTS

48 assays P-4027-48	96 assays P-4027-96
10 ml 6 ml 5 μl 12 μl	20 ml 12 ml 10 μl 24 μl
12μ l	24μ l
	8 ml
10μ l	20μ l
5μ l	10 <i>μ</i> l
120 <i>μ</i> Ι	240μ l
4	9
2	3
1	1
	P-4027-48 10 ml 6 ml 5 μl 12 μl 12 μl 4 ml 10 μl 5 μl 120 μl 4

^{*} For maximum recovery of the products, centrifuge the original vial prior to opening the cap.

SHIPPING & STORAGE

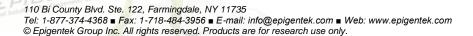
The kit is shipped in two parts, one part at ambient room temperature, and the second part on frozen ice packs at 4°C.

Upon receipt: (1) Store **F4** and **Standard Control** at –20°C; (2) Store **all other components** at 4°C away from light. The components of the kit are stable for 6 months from the date of shipment, when stored properly.

Note: Check if buffers **F1** and **F2** contain salt precipitates before using. If so, warm (at room temperature or 37°C) and shake the buffers until the salts are re-dissolved.

MATERIALS REQUIRED BUT NOT SUPPLIED

Orbital shaker
Pipettes and pipette tips
Reagent reservoir
Fluorescence microplate reade
15 ml conical tube
1.5 ml microcentrifuge tubes





GENERAL PRODUCT INFORMATION

Usage Limitation: The $EpiQuik^{TM}$ Global Acetyl Histone H4-K16 Quantification Kit (Fluorometric) is for research use only and is not intended for diagnostic or therapeutic application.

Safety: Suitable lab coat, disposable gloves, and eye protection are required when working with the kit.

Quality Control: Epigentek guarantees the performance of all products in the manner described in our product instructions.

Product Updates: Epigentek reserves the right to change or modify any product to enhance its performance and design.

Intellectual Property: The *EpiQuik*™ Global Acetyl Histone H4-K16 Quantification Kit (Fluorometric) and methods of use contain proprietary technologies by Epigentek. *EpiQuik*™ is a trademark of Epigentek Group Inc.

A BRIEF OVERVIEW

Acetylation of histones, including histone H3 and H4, has been involved in the regulation of chromatin structure and recruitment of transcription factors to the gene promoters. Histone acetyltransferases (HATs) and histone deacetylases (HDACs) play a critical role in the control of histone H4 acetylation at multiple sites. Acetylation of histone H4 at lysine 16 (H4-K16) reflects the hyperacetylated state in histone H4 and is strongly correlated with active states of genes. H4-K16 acetylation is related to euchromatin and DNA repair and hypoacetylation of H4-K16 is commonly found in human tumors. Histone H4-K16 acetylation may be increased by inhibition of HDACs and decreased by HAT inhibition; thus, quantitative detection of global acetyl histone H4-K16 would provide useful information for better understanding epigenetic regulation of gene activation and for developing HAT or HDAC-targeted drugs. The *EpiQuik*™ Global Acetyl Histone H4-K16 Quantification Kit (Fluorometric) provides a tool for measuring global acetylation of histone H4-K16. The kit has the following features:

- Quick and efficient procedure, which can be finished within 2.5 hours.
- Innovative fluorometric assay without the need for radioactivity, electrophoresis, or chromatography.
- Specifically captures acetyl H4-K16 with the detection limit as low as 0.4 ng/well and detection range from 20 ng-5 µg/well of histone extracts.
- The control is conveniently included for the quantification of the amount of acetyl H4-K16.
- Strip microplate format makes the assay flexible: manual or high throughput.
- Simple, reliable, and consistent assay conditions.

PRINCIPLE & PROCEDURE

The EpiQuik™ Global Acetyl Histone H4-K16 Quantification Kit (Fluorometric) is designed for measuring global histone H4-K16 acetylation. In an assay with this kit, the acetyl histone H4 at lysine 16 is captured to the strip wells coated with an anti-acetyl H4-K16 antibody. The captured



acetyl histone H4-K16 can then be detected with a labeled detection antibody, followed by a fluorescent development reagent. The ratio of acetyl H4-K16 is proportional to the intensity of fluorescence. The absolute amount of acetyl H4-K16 can be quantified by comparing to the standard control.



Schematic Procedure for the *EpiQuik™* Global Acetyl Histone H4-K16 Quantification Kit (Fluorometric)

PROTOCOL

- 1. **a)** Prepare histone extracts from cells/tissues treated or untreated by using your own successful method (acid extraction or high salt extraction).
 - **b)** For your convenience and the best results, Epigentek offers the *EpiQuik*™ Total Histone Extraction Kit (Cat. No. OP-0006) optimized for use in the *EpiQuik*™ modified histone quantification series.
 - c) Preparation of histone extracts can also be performed using the attached procedure. Histone extracts can be used immediately or stored at -80°C for future use.
- 2. Determine the number of strip wells required. Leave these strips in the plate frame (remaining unused strips can be placed back in the bag. Seal the bag tightly and store at 4°C). Dilute F1 with distilled water (pH 7.2-7.5) at a 1:9 ratio (1 ml of F1 + 9 ml of distilled water).
- 3. Add 50 μ l of **F2** into each well. For the sample, add 50-200 ng of the histone extract into the sample wells. For the standard curve, dilute the **Standard Control** with **F2** to 1 100 ng/ μ l at 5-7 points (e.g., 1.5, 3, 6, 12, 25, 50, and 100 ng/ μ l). Add 1 μ l of **Standard Control** at the different concentrations into the standard wells. For the blank, do not add any nuclear extracts or standard control protein. Mix and cover the strip wells with Parafilm M and incubate at room temperature for 1 hour.

Meanwhile, prepare the **detection solution**: for each 1 ml of **detection solution** to be prepared, add 1 μ l of **F3** and 0.5 μ l of **Signal Report Solution** into 10 μ l of **diluted F1**, mix and incubate at room temperature for 10 minutes. Next, add 20 μ l of **Signal Enhancer**, mix and incubate at room temperature for 15 minutes. Finally, add 970 μ l of **diluted F1** and mix.



- 4. Aspirate and wash the wells with 150 μ l of **diluted F1** three times.
- 5. Add 50 μ l of the **detection solution** to each well and incubate at room temperature for 60 minutes on an orbital shaker (100 rpm).
- 6. Aspirate and wash the wells with 150 μ l of **diluted F1** six times.
- 7. Prepare fluoro-development solution by adding 1 μ l of F4 and 1 μ l of F5 into each 400 μ l of F6. Add 50 μ l of fluoro-development solution into the wells and incubate at room temperature for 1-5 minutes away from light. The color in the standard wells containing the higher concentrations may turn slightly pink during this period. Measure and read fluorescence with a fluorescence microplate reader at $530_{\rm FX}/590_{\rm FM}$ nm.

Note: If the strip well frame does not fit in the fluorescence reader, transfer the solution to a standard 96-well microplate and read fluorescence at $530_{EX}/590_{EM}$ nm.

8. Calculate % histone H4-K16 acetylation:

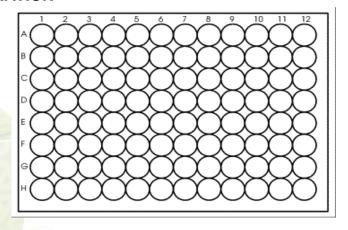
Acetylation % =
$$\frac{\text{RFU (treated (tested) sample - blank)}}{\text{RFU (untreated (control) sample - blank)}} \times 100\%$$

For the amount quantification, plot RFU versus amount of **Standard Control** and determine the slope as delta RFU/ng.

Calculate the amount of acetyl H4-K16 using the following formula:

Amount (ng/mg protein) =
$$\frac{\text{RFU (sample - blank)}}{\text{Protein } (\mu g)^* \times \text{slope}} \times 1000$$

PLATE CONFIGURATION



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^{*} Histone extract amount added into the sample well at step 3.



- Strip 1-3 (for 96 assays) or strip 1-2 (for 48 assays) standard wells (green trimmed); the standard curve can be generated with 5-8 concentration points (includes blank).
- Example amount of standard control/well A1: 100 ng; B1: 50 ng; C1: 25 ng; D1: 12 ng;
 E1: 6 ng; F1: 3 ng; G1: 1.5 ng; H1: 0 ng.
- Strip 4-12 (for 96 assays) or strip 3-6 (for 48 assays) sample wells (No label).
- Each sample or standard point can be assayed in duplicates or triplicates.

Histone Extraction Protocol

1. For tissues (treated and untreated), weigh the sample and cut the sample into small pieces (1-2 mm³) with a scalpel or scissors. Transfer tissue pieces to a Dounce homogenizer. Add TEB buffer (PBS containing 0.5% Triton X 100, 2 mM PMSF and 0.02% NaN₃) at 200 mg/ml, and disaggregate tissue pieces by 50-60 strokes. Transfer homogenized mixture to a 15 ml conical tube and centrifuge at 3000 rpm for 5 minutes at 4°C. If total mixture volume is less than 2 ml, transfer mixture to a 2 ml vial and centrifuge at 10,000 rpm for 1 minute at 4°C. Remove supernatant.

For cells (treated and untreated), harvest cells and pellet the cells by centrifugation at 1000 rpm for 5 minutes at 4° C. Resuspend cells in TEB buffer at 10^{7} cells/ml and lyse cells on ice for 10 minutes with gentle stirring. Centrifuge at 3000 rpm for 5 minutes at 4° C. If total volume is less than 2 ml, transfer cell lysates to a 2 ml vial and centrifuge at 10,000 rpm for 1 minute at 4° C. Remove supernatant.

- 2. Resuspend cell/tissue pellet in 3 volumes (approx. $200 \,\mu$ l/ 10^7 cells or 200 mg tissues) of extraction buffer (0.5N HCl + 10% glycerol) and incubate on ice for 30 min.
- 3. Centrifuge at 12,000 rpm for 5 minutes at 4°C and remove the supernatant fraction to new vial.
- 4. Add 8 volumes (approx. 0.6 ml/10⁷ cells or 200 mg tissues) of acetone and leave at -20°C overnight.
- 5. Centrifuge at 12,000 rpm for 5 minutes and air-dry the pellet. Dissolve the pellet in distilled water $(30-50 \,\mu\text{l}/10^7 \,\text{cells})$ or 200 mg tissues).
- 6. Quantify the protein concentration. Aliquot the extract and store the extract at -20°C or -80°C.

TROUBLESHOOTING

No Signal for Both the Standard Control and the Samples

Reagents are added incorrectly.

Check if reagents are added in the proper order and if any steps of the procedure are omitted by

mistake.

Incubation time and temperature are

Ensure the incubation time and temperature



incorrect.

described in the protocol are followed correctly.

No Signal or Very Weak Signal for Only the Standard Control

The amount of standard control is not added into "standard control wells" or is added insufficiently.

Ensure a sufficient amount of control is properly added to the standard control wells.

No Signal for Only the Sample

The protein sample is not properly extracted.

Ensure the procedure and reagents are correct for the nuclear protein extraction.

The protein amount is added into well insufficiently.

Ensure extract contains a sufficient amount of protein.

Protein extracts are stored incorrectly.

Ensure the protein extracts are stored at -20°C or -80°C.

High Background Present for the Blank

The well is not washed sufficiently.

Check if wash at each step is performed

according to the protocol.

Contaminated by the standard control.

Ensure the well is not contaminated from adding the control protein or by using control protein

contaminated tips.

Overdevelopment.

Decrease development time in step 7.

RELATED PRODUCTS

P-4022	EpiQuik™ Global Acetyl Histone H4-K5Quantification Kit (Colorimetric)
P-4023	EpiQuik™ Global Acetyl Histone H4-K5Quantification Kit (Fluorometric)
P-4024	EpiQuik™ Global Acetyl Histone H4-K8Quantification Kit (Colorimetric)
P-4025	EpiQuik™ Global Acetyl Histone H4-K8Quantification Kit (Fluorometric)
P-4026	<i>EpiQuik</i> ™ Global Acetyl Histone H4-K16Quantification Kit (Colorimetric)
P-4028	<i>EpiQuik</i> ™ Global Acetyl Histone H4-K12Quantification Kit (Colorimetric)
P-4029	EpiQuik [™] Global Acetyl Histone H4-K12 Quantification Kit (Fluorometric)

