EpiQuik™ Global Tri-Methyl Histone H3-K36 Quantification Kit (Fluorometric)

Base Catalog # P-3051

PLEASE READ THIS ENTIRE USER GUIDE BEFORE USE

The EpiQuik™ Global Tri-Methyl Histone H3-K36 Quantification Kit (Fluorometric) is suitable for specifically measuring global histone H3-K36 tri-methylation using a variety of mammalian cells (human, mouse, etc.) including fresh and frozen tissues, cultured adherent and suspension cells.
KIT CONTENTS

<table>
<thead>
<tr>
<th>Components</th>
<th>48 assays</th>
<th>96 assays</th>
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</thead>
<tbody>
<tr>
<td>F1 (10X Wash Buffer)</td>
<td>10 ml</td>
<td>20 ml</td>
</tr>
<tr>
<td>F2 (Antibody Buffer)</td>
<td>6 ml</td>
<td>12 ml</td>
</tr>
<tr>
<td>F3 (Detection Antibody, 1 mg/ml)*</td>
<td>5 µl</td>
<td>10 µl</td>
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<tr>
<td>F4 (Fluoro Developer)*</td>
<td>12 µl</td>
<td>24 µl</td>
</tr>
<tr>
<td>F5 (Fluoro Enhancer)*</td>
<td>12 µl</td>
<td>24 µl</td>
</tr>
<tr>
<td>F6 (Fluoro Dilution)</td>
<td>4 ml</td>
<td>8 ml</td>
</tr>
<tr>
<td>Standard Control (100 µg/ml)*</td>
<td>10 µl</td>
<td>20 µl</td>
</tr>
<tr>
<td>8-Well Sample Strips (with Frame)</td>
<td>4</td>
<td>9</td>
</tr>
<tr>
<td>8-Well Standard Control Strips</td>
<td>2</td>
<td>3</td>
</tr>
</tbody>
</table>

* For maximum recovery of the products, centrifuge the original vial prior to opening the cap.

SHIPPING & STORAGE

Upon receipt, store F3, F4 and Standard control at –20°C. Store all other components at 4°C away from light. The components of the kit should be stable for 6 months when stored properly.

**Note:** Check if buffers F1 and F2 contain salt precipitates before using. If so, warm (at room temperature or 37°C) and shake the buffers until the salts are re-dissolved.

MATERIALS REQUIRED BUT NOT SUPPLIED

- Orbital shaker
- Pipettes and pipette tips
- Reagent reservoir
- Fluorescence microplate reader
- 15 ml conical tube
- 1.5 ml microcentrifuge tubes

GENERAL PRODUCT INFORMATION

**Usage Limitation:** The EpiQuik™ Global Tri-Methyl Histone H3-K36 Quantification Kit (Fluorometric) is for research use only and is not intended for diagnostic or therapeutic application.

**Safety:** Suitable lab coat, disposable gloves, and eye protection are required when working with the kit.

**Quality Control:** EpigenTek guarantees the performance of all products in the manner described in our product instructions.
Product Updates: EpigenTek reserves the right to change or modify any product to enhance its performance and design.

Intellectual Property: The EpiQuik™ Global Tri-Methyl Histone H3-K36 Quantification Kit (Fluorometric) and methods of use contain proprietary technologies by Epigentek. EpiQuik™ is a trademark of EpigenTek Group Inc.

A BRIEF OVERVIEW

Epigenetic activation or inactivation of genes plays a critical role in many important human diseases, especially in cancer. A major mechanism for epigenetic inactivation of the genes is methylation of CpG islands in genome DNA caused by DNA methyltransferases. Histone methyltransferases (HMTs) control or regulate DNA methylation through chromatin-dependent transcription repression or activation. HMTs transfer 1-3 methyl groups from S-adenosyl-L-methionine to the lysine and arginine residues of histone proteins. SET2 is a histone methyltransferase that catalyzes methylation of histone H3 at lysine 36 (H3-K36) in mammalian cells. H3-K36 tri-methylation is associated with transcriptionally active genes. Increased global H3-K36 methylation is also found to be linked to the Sotos syndrome and leukemia-associated protein NSD1 and the Huntington’s disease protein HYPB. The global H3-K36 tri-methylation can be changed by inhibition or activation of HMTs. Thus, quantitative detection of global di-methyl histone H3-K4 would provide useful information for better understanding epigenetic regulation of gene activation and for developing HMT-targeted drugs. The EpiQuik™ Global Tri-Methyl Histone H3-K36 Quantification Kit (Fluorometric) provides a tool for measuring global tri-methylation of histone H3-K36. The kit has the following features:

- Quick and efficient procedure, which can be finished within 2.5 hours.
- Innovative fluorometric assay with no need for radioactivity, electrophoresis, and chromatography.
- Specifically capturing tri-methylated H3-K36 with the detection limit as low as 0.4 ng/well and detection range from 5 ng-2 µg/well of histone extracts.
- The control is conveniently included for quantification of the amount of tri-methylated H3-K36.
- Strip microplate format makes the assay flexible: manual or high throughput.
- Simple, reliable, and consistent assay conditions

PRINCIPLE & PROCEDURE

The EpiQuik™ Global Tri-Methyl Histone H3-K36 Quantification Kit (Fluorometric) is designed for measuring global histone H3-K36 tri-methylation. In an assay with this kit, the tri-methylated histone H3 at lysine 36 is captured to the strip wells coated with anti-trimethyl H3-K36 antibody. The captured tri-methylated histone H3-K36 can be then detected with a labeled detection antibody followed by fluorescent development reagent. The ratio of tri-methylated H3-K36 is proportional to the intensity of fluorescence. The absolute amount of tri-methylated H3-K36 can be quantified by comparing to the standard control.
PROTOCOL

1. **a)** Prepare histone extracts from cells/tissues treated or untreated by using your own successful method (acid extraction or high salt extraction).
   **b)** For your convenience and the best results, EpigenTek offers the EpiQuik™ Total Histone Extraction kit (Cat # OP-0006) optimized for use in the EpiQuik™ modified histone quantification series.
   **c)** Preparation of histone extracts can also be performed using the attached procedure. Histone extracts can be used immediately or stored at –80°C for future use.

2. Determine the number of the strip wells required. Leave these strips in the plate frame (remaining unused strips can be put back in the bag. Seal the bag tightly and store at 4°C). Dilute F1 with distilled water (pH 7.2-7.5) at a 1:9 ratio (1 ml of F1 + 9 ml of distilled water).

3. Add 50 µl of F2 into each well. For the sample, add 50-200 ng of the histone extract into the sample wells. For standard curve, dilute standard control with F2 to 1 – 100 ng/µl for 5-7 points (ie: 1.5, 3, 6, 12, 25, 50, and 100 ng/µl). Add 1 µl of standard control at the different concentrations into the standard well. For the blank, add no nuclear extracts or no standard control protein. Mix and cover the strip wells with Parafilm M and incubate at room temperature for 1-2 hours.

4. Aspirate and wash the wells with 150 µl of diluted F1 3 times.

5. Dilute F3 (at 1:1000 ratio) to 1 µg/ml with F2. Add 50 µl of diluted F3 to each well and incubate at room temperature for 60 min on an orbital shaker (100 rpm).

6. Aspirate and wash the wells with 150 µl of diluted F1 6 times.

7. Prepare fluoro-development solution by adding 1 µl of F4 and 1 µl of F5 into each 400 µl of F6. Add 50 µl of fluoro development solution into the wells and incubate at room temperature for 1-5 min away from light. The color in the standard wells containing the higher concentrations may turn slightly pink during this period. Measure and read fluorescence on fluorescence microplate reader at 530EX/590EM nm.
Note: If the strip well frame does not fit the fluorescence reader, transfer the solution to a standard 96-well microplate and read fluorescence at 530<sub>EX</sub>/590<sub>EM</sub> nm.

8. Calculate % histone H3-K36 tri-methylation:

\[
\text{Tri-methylation %} = \frac{\text{RFU (treated (tested) sample – blank)}}{\text{RFU (untreated (control) sample – blank)}} \times 100\%
\]

For the amount quantification, plot RFU versus amount of standard control and determine the slope as delta RFU/ng.

Calculate the amount of tri-methylated H3-K36 using the following formula:

\[
\text{Amount (ng/mg protein)} = \frac{\text{RFU (sample – blank)}}{\text{Protein (µg)}} \times \text{slope} \times 1000
\]

* Histone extract amount added into the sample well at step 3

**PLATE CONFIGURATION**

- Strip 1-3 (for 96 assays) or strip 1-2 (for 48 assays): standard wells (green trimmed): The standard curve can be generated with 5-8 concentration points (includes blank).
- Example amount of standard control /well: A1: 100 ng; B1: 50 ng; C1: 25 ng; D1: 12 ng; E1: 6 ng; F1: 3 ng; G1 1.5 ng; H1: 0 ng
- Strip 4-12 (for 96 assays) or strip 3-6 (for 48 assays): sample wells (No label)
- Each sample or standard point can be assayed in the duplicates or triplicates.
Appendix

Histone Extraction Protocol

1. For tissues (treated and untreated). Weigh the sample and cut the sample into small pieces (1-2 mm³) with a scalpel or scissors. Transfer tissue pieces to a Dounce homogenizer, add TEB buffer (PBS containing 0.5% Triton X 100, 2 mM PMSF and 0.02% NaN₃) at 200 mg/ml, and disaggregate tissue pieces by 50-60 strokes. Transfer homogenized mixture to a 15 ml conical tube and centrifuge at 3,000 rpm for 5 min at 4°C. If total mixture volume is less than 2 ml, transfer mixture to a 2 ml vial and centrifuge at 10,000 rpm for 1 min at 4°C. Remove supernatant.

For cells (treated and untreated). Harvest cells and pellet the cells by centrifugation at 1000 rpm for 5 min at 4°C. Resuspend cells in TEB buffer at 10⁷ cells/ml and lyse cells on ice for 10 min with gentle stirring. Centrifuge at 3000 rpm for 5 min at 4°C. If total volume is less than 2 ml, transfer cell lysates to a 2 ml vial and centrifuge at 10000 rpm for 1 min at 4°C. Remove supernatant.

2. Resuspend cell/tissue pellet in 3 volumes (approx. 200 µl/10⁷ cells or 200 mg tissues) of extraction buffer (0.5N HCl + 10% glycerol) and incubate on ice for 30 min.

3. Centrifuge at 12,000 rpm for 5 min at 4°C and remove the supernatant fraction to new vial.

4. Add 8 volumes (approx. 0.6 ml/10⁷ cells or 200 mg tissues) of acetone and leave at −20°C overnight.

5. Centrifuge at 12,000 rpm for 5 min and air-dry the pellet. Dissolve the pellet in distilled water (30-50 µl/10⁷ cells or 200 mg tissues).

6. Quantify the protein concentration. Aliquot the extract and store the extract at −20°C or −80°C.

TROUBLESHOOTING

No Signal for Both the Standard Control and the Samples

Reagents are added incorrectly. Check if reagents are added in order and if some steps of the procedure are omitted by mistake.

Incubation time and temperature are incorrect. Ensure the incubation time and temperature described in the protocol are correctly followed.

No Signal or Very Weak Signal for Only the Standard Control

The amount of standard control is not added into “standard control wells” or is added insufficiently Ensure sufficient amount of control is properly added to the standard control well.
No Signal for Only the Sample

The protein sample is not properly extracted. Ensure the procedure and reagents are correct for the nuclear protein extraction.

The protein amount is added into well insufficiently. Ensure extract contains sufficient amount of proteins.

Protein extracts are incorrectly stored. Ensure the nuclear extracts are stored at −20°C or −80°C.

High Background Present for the Blank

The well is not washed enough. Check if wash at each step is performed according to the protocol.

Contaminated by the standard control. Ensure the well is not contaminated by adding the control protein or by using control protein contaminated tips.

Overdevelopment. Decrease color development time in Step 7

RELATED PRODUCTS

P-3046    EpiQuik™ Global Mono-Methyl Histone H3-K36 Quantification Kit (Colorimetric)
P-3047    EpiQuik™ Global Mono-Methyl Histone H3-K36 Quantification Kit (Fluorometric)
P-3048    EpiQuik™ Global Di-Methyl Histone H3-K36 Quantification Kit (Colorimetric)
P-3049    EpiQuik™ Global Di-Methyl Histone H3-K36 Quantification Kit (Fluorometric)
P-3050    EpiQuik™ Global Tri-Methyl Histone H3-K36 Quantification Kit (Colorimetric)
P-3052    EpiQuik™ Global Pan-Methyl Histone H3-K36 Quantification Kit (Colorimetric)
P-3053    EpiQuik™ Global Pan-Methyl Histone H3-K36 Quantification Kit (Fluorometric)