

# EpiQuik™ 8-OHdG DNA Damage Quantification Direct Kit (Fluorometric)

Base Catalog # P-6004

## PLEASE READ THIS ENTIRE USER GUIDE BEFORE USE

**Uses:** The EpiQuik™ 8-OHdG DNA Damage Quantification Direct Kit (Fluorometric) is suitable for detecting oxidative DNA damage (8-OHdG) status using DNA isolated from any species such as mammals, plants, fungi, bacteria, and viruses in a variety of forms including, but not limited to, cultured cells, fresh and frozen tissues, paraffin-embedded tissues, and body fluid samples.

**Input DNA:** The amount of DNA for each assay can be 100 ng to 300 ng. For optimal quantification, the input DNA amount should be 300 ng, as basal 8-OHdG is generally less than 0.01% of total DNA.

**Starting Material:** Starting materials can include various tissue or cell samples such as cells from flask or microplate cultured cells, fresh and frozen tissues, paraffin-embedded tissues, blood, body fluid samples, etc.

**Internal Control:** Both negative and positive DNA controls are provided in this kit. A standard curve can be performed (range: 5 to 200 pg of 8-OHdG) or a single quantity of 8-OHdG can be used as a positive control. Because 8-OHdG content can vary from tissue to tissue, and from normal and diseased states, or vary under treated and untreated conditions, it is advised to run replicate samples to ensure that the signal generated is validated. This kit will allow the user to quantify an absolute amount of 8-OHdG and determine the relative 8-OHdG states of two different DNA samples.

**Precautions:** To avoid cross-contamination, carefully pipette the sample or solution into the strip wells. Use aerosol-barrier pipette tips and always change pipette tips between liquid transfers. Wear gloves throughout the entire procedure. In case of contact between gloves and sample, change gloves immediately.

## KIT CONTENTS

Component	48 Assays Cat. #P-6004-48	96 Assays Cat. #P-6004-96	Storage Upon Receipt
<b>WB</b> (10X Wash Buffer)	14 ml	28 ml	4°C
<b>BS</b> (Binding Solution)	5 ml	10 ml	RT
<b>NC</b> (Negative Control , 10 µg/ml)*	10 µl	20 µl	-20°C
<b>PC</b> (Positive Control, 8-OHdG 0.2 µg/ml)*	10 µl	20 µl	-20°C
<b>CA</b> (Capture Antibody, 500 X) *	8 µl	16 µl	4°C
<b>DA</b> (Detection Antibody, 1000 X)*	6 µl	12 µl	-20°C
<b>ES</b> (Enhancer Solution)*	5 µl	10 µl	-20°C
<b>FD</b> (Fluoro-Developer)*	10 µl	20 µl	-20°C
<b>FE</b> (Fluoro-Enhancer)*	10 µl	20 µl	4°C
<b>DB</b> (Dilution Buffer)	4 ml	8 ml	RT
8-Well Assay Strips (With Frame)	6	12	4°C

\* Spin the solution down to the bottom prior to use.

**Note:** The **NC** Negative Control I is an oligos containing no 8-OHdG. The **PC** Positive Control is an 8-OHdG oligos and normalized to have 100% of 8-OHdG.

## SHIPPING & STORAGE

The kit is shipped in two parts: the first part at ambient room temperature and the second part on frozen ice packs at 4°C.

Upon receipt: (1) Store **NC**, **PC**, **DA**, **ES** and **FD** at -20°C away from light; (2) Store **WB**, **CA**, **DS**, **FE** and **8-Well Assay Strips** at 4°C away from light; (3) Store remaining components (**BS** and **DB**) at room temperature away from light.

**Note:** Check if wash buffer, **WB**, contains salt precipitates before using. If so, warm (at room temperature or 37°C) and shake the buffer until the salts are re-dissolved.

All components of the kit are stable for 6 months from the date of shipment, when stored properly.

## MATERIALS REQUIRED BUT NOT SUPPLIED

- Adjustable pipette
- Aerosol resistant pipette tips
- Microplate reader capable of reading fluorescence 530ex/590em
- 1.5 ml microcentrifuge tubes
- Incubator for 37°C incubation
- Plate seal or Parafilm M

- Distilled water
- 1X TE buffer pH 7.5 to 8.0
- Isolated DNA of interest

## GENERAL PRODUCT INFORMATION

**Quality Control:** Each lot of the EpiQuik™ 8-OHdG DNA Damage Quantification Direct Kit (Fluorometric) is tested against predetermined specifications to ensure consistent product quality. EpigenTek guarantees the performance of all products in the manner described in our product instructions.

**Product Warranty:** If this product does not meet your expectations, simply contact our technical support unit or your regional distributor. We also encourage you to contact us if you have any suggestions about product performance or new applications and techniques.

**Safety:** Suitable lab coat, disposable gloves, and proper eye protection are required when working with this product.

**Product Updates:** EpigenTek reserves the right to change or modify any product to enhance its performance and design. The information in this User Guide is subject to change at any time without notice. Be sure to use the latest User Guide for this kit which can be accessed online at [www.epigentek.com/datasheet](http://www.epigentek.com/datasheet).

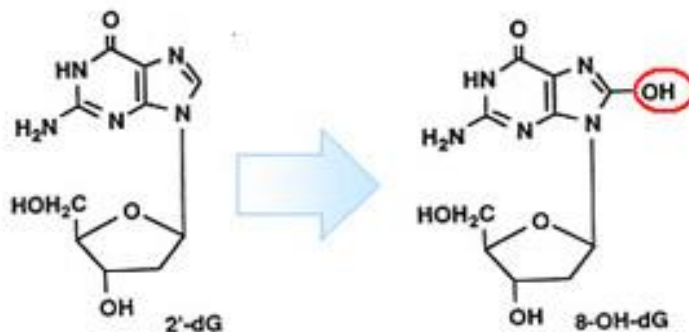
**Usage Limitation:** The EpiQuik™ 8-OHdG DNA Damage Quantification Direct Kit (Fluorometric) is for research use only and is not intended for diagnostic or therapeutic application.

**Intellectual Property:** The EpiQuik™ 8-OHdG DNA Damage Quantification Direct Kit (Fluorometric) and methods of use contain proprietary technologies by EpigenTek.

## A BRIEF OVERVIEW

8-hydroxy-2'-deoxyguanosine (8-OHdG or 8-oxo-dG) is an oxidized derivative of deoxyguanosine and is generated by hydroxyl radicals, singlet oxygen, and one-electron oxidants in cellular DNA. As a modified nucleoside base, 8-OHdG is considered important not only because of its abundance but also because of its mutagenic potential through G-to-T transversion mutations upon replication of DNA. 8-OHdG also participates in epigenetic regulation of gene activation/repression by inhibiting the binding affinity of MBD proteins to the CpG sites of DNA. It is generally accepted that oxidatively damaged DNA can be repaired by base excision repair (BER) enzymes.

Currently, 8-OHdG is widely accepted as a sensitive marker of oxidative DNA damage and oxidative stress. Evidence shows that increased levels of 8-OHdG are closely correlated with exposure to harmful environmental factors such as ionizing radiation, industrial chemicals, air pollution, cigarette smoking, and cancer chemotherapy. It has also been demonstrated that increased concentrations of 8-OHdG are pathogenically linked to a variety of age-associated diseases including cancer, coronary heart disease, diabetes and neurodegenerative diseases such as Alzheimer's disease. Compared with the urine 8-OHdG assay that mainly reflects the balance between oxidative damage and the repair rate of the whole body, directly quantifying the 8-OHdG content in different cells/tissues in normal and disease states would allow tissue-specific oxidative damage of DNA to be identified. Therefore, more useful information for the better understanding of oxidative damage-disease relationships, which benefits diagnostics and therapeutics of the disease, can be obtained.



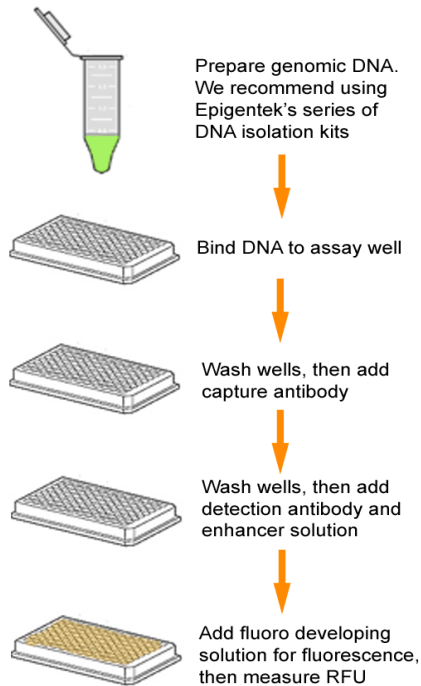
Formation of 8-hydroxy-2'-deoxyguanosine (8-OHdG) by oxygen radicals. H. Kasai:  
 Environmental Mutagen Research, Vol. 10, p73-78 (1988)

Several chromatography-based techniques such as HPLC-ECD and LC-MS are used for detecting 8-OHdG in tissues and cells. However, these methods are time consuming and have low throughput with high costs. The currently used competitive ELISA methods are also not conveniently applicable for cell/tissue 8-OHdG detection because they are less accurate and have an inability to use intact DNA isolated from cells or tissues directly. To address these problems, EpigenTek offers the EpiQuik™ 8-OHdG DNA Damage Quantification Direct Kit (Fluorometric) which uses a unique procedure to directly quantify 8-OHdG in cells/tissues. The kit has the following advantages and features:

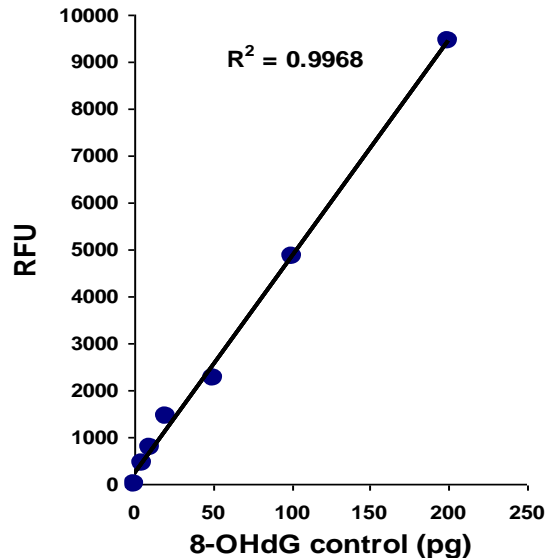
- Fluorometric assay with easy-to-follow steps for convenience and speed. The entire procedure can be completed within 3 hours and 45 minutes.
- High sensitivity, of which the detection limit can be as low as 1 pg of 8-OHdG.
- High specificity by detecting only 8-OHdG without cross-reactivity to 8-OHdG analogues such as dG, guanine, 8-OHGua and 8-OHG within the indicated concentration range of the sample DNA.
- Direct detection of 8-OHdG using intact DNA, which eliminates interference from high molecular weight compounds such as carbohydrates and proteins that are often seen in competitive 8-OHdG assays.
- Detection accuracy is highly correlated with and close to HPLC or LC-MS analysis.
- Highly convenient assay with direct use of DNA isolated from cells or tissues, no need for DNA digestion or hydrolysis.
- Universal positive and negative controls are included, which are suitable for quantifying 8-OHdG from any species.
- Strip-well microplate format makes the assay flexible for manual or high throughput analysis.
- Simple, reliable, and consistent assay conditions

## PRINCIPLE & PROCEDURE

The EpiQuik™ 8-OHdG DNA Damage Quantification Direct Kit (Fluorometric) contains all reagents necessary for the quantification of Oxidative DNA damage (8-OHdG). In this assay, DNA is bound to strip wells that are specifically treated to have a high DNA affinity. 8-OHdG is detected using capture and detection antibodies. The detected signal is enhanced and then quantified fluorometrically by reading the fluorescence in a fluorescence microplate reader. The amount of 8-OHdG is proportional to the fluorescence intensity measured.



Schematic procedure of the EpiQuik™ 8-OHdG DNA Damage Quantification Direct Kit (Fluorometric)



8-OHdG standard control was added into the assay wells at different concentrations and then measured with the EpiQuik™ 8-OHdG DNA Damage Quantification Direct Kit (Fluorometric).

## ASSAY PROTOCOL

For the best results, please read the protocol in its entirety prior to starting your experiment.

### 1. Starting Materials

**Input DNA Amount:** DNA amount can range from 100 ng to 300 ng per reaction. An optimal amount is 300 ng per reaction. Starting DNA may be in water or in a buffer such as TE.

**DNA Isolation:** You can use your method of choice for DNA isolation. EpigenTek offers a series of genomic DNA isolation kits for your convenience (see "Related Products" section).

**DNA Storage:** Isolated genomic DNA can be stored at 4°C (short term) or -20°C (long term) until use.

### 2. Buffer and Solution Preparation

- a. Preparation of 1X Wash Buffer:

**48-Assay Kit:** Add 13 ml of **WB** 10X Wash Buffer to 117 ml of distilled water (pH 7.2-7.5).

**96-Assay Kit:** Add 26 ml of **WB** 10X Wash Buffer to 234 ml of distilled water (pH 7.2-7.5).

This **Diluted WB** 1X Wash Buffer can now be stored at 4°C for up to six months.

- b. Prepare Diluted **CA** Capture Antibody Solution:

Dilute **CA** (Capture Antibody) with **Diluted WB** at a ratio of 1:500 (i.e., add 1  $\mu$ l of **CA** to 500  $\mu$ l of **Diluted WB**). About 50  $\mu$ l of this **Diluted CA** will be required for each assay well.

- c. Prepare Diluted **DA** Detection Antibody Solution:

Dilute **DA** (Detection Antibody) with **Diluted WB1** at a ratio of 1:1000 (i.e., add 1  $\mu$ l of **DA** to 1000  $\mu$ l of **Diluted WB**). About 50  $\mu$ l of this **Diluted DA** will be required for each assay well.

- d. Prepare Diluted **ES** Enhancer Solution:

Dilute **ES** Enhancer Solution with **Diluted WB** at a ratio of 1:5000 (i.e., add 1  $\mu$ l of **ES** to 5000  $\mu$ l of **Diluted WB**). About 50  $\mu$ l of this **Diluted ES** will be required for each assay well.

- e. Prepare Fluorescence Development Solution:

Add 1  $\mu$ l of **FD** (Fluoro Developer) and 1  $\mu$ l of **FE** (Fluoro Enhancer) to every 500  $\mu$ l of **DB** (Dilution Buffer). About 50  $\mu$ l of this **Fluorescence Development Solution** will be required for each assay well

- f. Preparation of Diluted Positive Control:

Single Point Control Prep: Dilute **PC** Positive Control with 1X TE to 100 pg/ $\mu$ l (1  $\mu$ l **PC** + 1  $\mu$ l TE).

Suggested Standard Curve Prep Prepare 6 different concentrations with the stock **PC** and 1X TE into 5, 10, 20, 50, 100 and 200 pg/ $\mu$ l according to the following dilution chart:

Tube	PC (200 pg/ $\mu$ l)	1X TE	Resulting PC Concentration
1	1.0 $\mu$ l	39.0 $\mu$ l	5 pg/ $\mu$ l
2	1.0 $\mu$ l	19.0 $\mu$ l	10 pg/ $\mu$ l
3	1.0 $\mu$ l	9.0 $\mu$ l	20 pg/ $\mu$ l
4	1.0 $\mu$ l	3.0 $\mu$ l	50 pg/ $\mu$ l
5	2.0 $\mu$ l	2.0 $\mu$ l	100 pg/ $\mu$ l
6	3.0 $\mu$ l	0.0 $\mu$ l	200 pg/ $\mu$ l

**Note:** Keep each of the diluted solutions (except **Diluted WB** 1X Wash Buffer) on ice until use. Any remaining diluted solutions, other than **Diluted WB**, should be discarded if not used within the same day.

### 3. DNA Binding

- Predetermine the number of strip wells required for your experiment. Carefully remove un-needed strip wells from the plate frame and place them back in the bag (seal the bag tightly and store at 4°C).
- Add 80  $\mu$ l of **BS** Binding Solution to each well.
- Add 1  $\mu$ l of **NC**, 1  $\mu$ l of **Diluted PC** (see note below), and 300 ng of your Sample DNA (1-8  $\mu$ l) into the designated wells depicted in [Table 1](#) or [Table 2](#). Mix solution by gently tilting from side to side or shaking the plate several times. Ensure the solution coats the bottom of the well evenly.

**Note:** (1) For a single point control, add 1  $\mu$ l of **PC** at a concentration of 100 pg/ $\mu$ l as prepared in Step 2f; For the standard curve, add 1  $\mu$ l of **Diluted PC** at concentrations of 5 to 200 pg/ $\mu$ l (see the chart in Step 2). The final amounts should be 5, 10, 20, 50, 100 and 200 pg per well. (2) For optimal binding, sample DNA volume added should not exceed 8  $\mu$ l.

- Cover strip plate with plate seal or Parafilm M and incubate at 37°C for 90 min.
- Remove the **BS** Binding Solution from each well. Wash each well with 150  $\mu$ l of the **Diluted WB** 1X Wash Buffer each time for three times.

#### **4. 8-OHdG DNA Capture**

- a. Add 50 µl of the **Diluted CA** to each well, then cover and incubate at room temperature for 60 min.
- b. Remove the **Diluted CA** solution from each well.
- c. Wash each well with 150 µl of the **Diluted WB** each time for three times.
- d. Add 50 µl of the **Diluted DA** to each well, then cover and incubate at room temperature for 30 min.
- e. Remove the **Diluted DA** solution from each well.
- f. Wash each well with 150 µl of the **Diluted WB** each time for four times.
- g. Add 50 µl of the **Diluted ES** to each well, then cover and incubate at room temperature for 30 min.
- h. Remove the **Diluted ES** solution from each well.
- i. Wash each well with 150 µl of the **Diluted WB** each time for five times.

#### **5. Signal Detection**

- a. Add 50 µl of **Fluorescence Development Solution** to each well and incubate at room temperature for 2 to 4 min away from direct light. The **Fluorescence Development Solution** will turn pink in the presence of sufficient 8-OHdG products.
- b. Read the fluorescence on a fluorescence microplate reader within 2 to 10 min at 530<sub>ex</sub>/590<sub>em</sub> nm.

**Note:** If the strip-well microplate frame does not fit in the fluorescence microplate reader, transfer the solution to a standard 96-well microplate.

#### **6. 8-OHdG Calculation**

**Relative Quantification:** To determine the relative 8-OHdG status of two different DNA samples, a simple calculation for the percentage of 8-OHdG in your total DNA can be carried out using the following formula:

$$8\text{-OHdG \%} = \frac{(\text{Sample RFU} - \text{NC RFU}) \div S}{(\text{PC RFU} - \text{NC RFU}) \div P} \times 100\%$$

**S** is the amount of input sample DNA in ng.

**P** is the amount of input positive control (**PC**) in ng.

Example calculation:

Average RFU of NC is 1500  
 Average RFU of PC is 5500  
 Average RFU of Sample is 2100  
 S is 300 ng  
 P is 0.1 ng (100 pg)

$$8\text{-OHdG \%} = \frac{(2100 - 1500) \div 300}{(5500 - 1500) \div 0.1} \times 100\% = 0.005\%$$

**Absolute Quantification:** To quantify the absolute amount of 8-OHdG using an accurate calculation, first generate a standard curve and plot the OD values versus the amount of **PC** at each concentration point. Next, determine the slope (OD/ng) of the standard curve using linear regression (*Microsoft Excel's* linear regression functions are suitable for such calculation) and also determine the most linear

part (include at least 4 concentration points) of the standard curve for optimal slope calculation. Now calculate the amount and percentage of 8-OHdG in your total DNA using the following formulas:

$$8\text{-OHdG (ng)} = \frac{\text{Sample RFU} - \text{NC RFU}}{\text{Slope}}$$

$$8\text{-OHdG \%} = \frac{8\text{-OHdG Amount}}{S} \times 100\%$$

**S** is the amount of input sample DNA in ng.

Example calculation:

Average RFU of NC is 1500  
 Average RFU of sample is 2100  
 Slope is 40000 RFU/ng  
 S is 300 ng

$$8\text{-OHdG (ng)} = \frac{2100 - 1500}{40000} = 0.015 \text{ ng}$$

$$8\text{-OHdG \%} = \frac{0.015}{300} \times 100\% = 0.005\%$$

## SUGGESTED STRIP WELL SETUP

**Table 1.** The suggested strip-well plate setup using a single point positive control in a 48-assay format (in a 96-assay format, Strips 7 to 12 can be configured as Sample). The controls and samples can be measured in duplicate.

Well #	Strip 1	Strip 2	Strip 3	Strip 4	Strip 5	Strip 6
A	NC	NC	Sample	Sample	Sample	Sample
B	PC	PC	Sample	Sample	Sample	Sample
C	Sample	Sample	Sample	Sample	Sample	Sample
D	Sample	Sample	Sample	Sample	Sample	Sample
E	Sample	Sample	Sample	Sample	Sample	Sample
F	Sample	Sample	Sample	Sample	Sample	Sample
G	Sample	Sample	Sample	Sample	Sample	Sample
H	Sample	Sample	Sample	Sample	Sample	Sample

**Table 2.** The suggested strip-well plate setup for standard curve preparation in a 48-assay format (in a 96-assay format, Strips 7 to 12 can be configured as Sample). The controls and samples can be measured in duplicate.

Well #	Strip 1	Strip 2	Strip 3	Strip 4	Strip 5	Strip 6
A	NC	NC	Sample	Sample	Sample	Sample
B	PC 5 pg/μl	PC 5 pg/μl	Sample	Sample	Sample	Sample
C	PC10 pg/μl	PC10 pg/μl	Sample	Sample	Sample	Sample
D	PC 20 pg/μl	PC 20 pg/μl	Sample	Sample	Sample	Sample
E	PC 50 pg/μl	PC 50 pg/μl	Sample	Sample	Sample	Sample
F	PC 100 pg/μl	PC 100 pg/μl	Sample	Sample	Sample	Sample
G	PC 200 pg/μl	PC 200 pg/μl	Sample	Sample	Sample	Sample
H	Sample	Sample	Sample	Sample	Sample	Sample



## SUGGESTED WORKING BUFFER AND SOLUTION SETUP

**Table 3.** Approximate amount of required buffers and solutions for defined assay wells based on the protocol.

Reagents	1 well	8 wells (1 strip)	16 wells (2 strips)	48 wells (6 strips)	96 wells (12 strips)
<b>Diluted WB</b>	2.5 ml	20 ml	40 ml	120 ml	240 ml
<b>BS</b>	80 $\mu$ l	640 $\mu$ l	1300 $\mu$ l	3900 $\mu$ l	8000 $\mu$ l
<b>Diluted CA</b>	50 $\mu$ l	400 $\mu$ l	800 $\mu$ l	2400 $\mu$ l	4800 $\mu$ l
<b>Diluted DA</b>	50 $\mu$ l	400 $\mu$ l	800 $\mu$ l	2400 $\mu$ l	4800 $\mu$ l
<b>Diluted ES</b>	50 $\mu$ l	400 $\mu$ l	800 $\mu$ l	2400 $\mu$ l	4800 $\mu$ l
<b>DS</b>	0.1 ml	0.8 ml	1.6 ml	4.8 ml	9.6 ml
<b>SS</b>	0.1 ml	0.8 ml	1.6 ml	4.8 ml	9.6 ml
<b>NC</b>	N/A	0.5 $\mu$ l – 1 $\mu$ l	0.5 $\mu$ l – 2 $\mu$ l	1 $\mu$ l – 4 $\mu$ l	2 $\mu$ l – 8 $\mu$ l
<b>PC</b>	N/A	0.5 $\mu$ l – 1 $\mu$ l	0.5 $\mu$ l – 2 $\mu$ l	1 $\mu$ l – 4 $\mu$ l	2 $\mu$ l – 8 $\mu$ l

## TROUBLESHOOTING

Problem	Possible Cause	Suggestion
No signal in both the positive control and sample wells	Reagents are added incorrectly.	Check if reagents are added in the proper order and if any steps in the protocol may have been omitted by mistake.
	The well is incorrectly washed before DNA binding.	Ensure the well is not washed prior to adding the positive control and sample.
	<b>BS</b> Binding Solution does not completely cover the well bottoms.	Ensure the solution coats the bottom of the well by gently tilting from side to side or shaking the plate several times.
	Incorrect time and temperature used.	Ensure the incubation time and temperature described in the protocol is followed correctly.
	Insufficient input materials.	Ensure that a sufficient amount of positive control (> 50 pg) and sample (300 ng) is added into the wells.
	Incorrect fluorescence reading.	Check if fluorescence filters (530ex/590em) are used.
	Kit was not stored or handled properly.	Ensure all components were stored at the appropriate temperature and the cap is closed tightly after each opening.
No signal or weak signal in only the positive control wells	Insufficient positive control added to the well in Step 3c.	Ensure a sufficient amount of positive control DNA is added.
	The <b>PC</b> Positive Control is degraded due to improper storage conditions.	Follow the Shipping & Storage guidelines of this User Guide for storage of <b>PC</b> Positive Control.
High background present in the negative control wells or RFU in some of	Insufficient washing of wells.	Check if washing recommendations at each step are performed according to the protocol.
	Contaminated by sample or positive control.	Ensure the well is not contaminated by the sample or positive control DNA or from the use of contaminated tips.

standard and sample wells is out of the fluorescent reading range.	Incubation time is too long.	The incubation time should not exceed 2 h at Step 3d, and not exceed 45 min at Step 4d.
	Over development of fluorescence.	Decrease the development time in Step 4a. Also, RFU can be re-measured after diluting the already fluorescent solution in the wells with PBS at 1:5 or 1:10 ratio. The RFU ratio remains unchanged in the designated assay wells.
	DNA sample is not properly extracted or purified.	Ensure the DNA sample is in good quality. 260/280 ratio should be >1.7 with no or minimal RNA contamination
No signal or weak signal only in sample wells	Sample amount added into the wells is insufficient.	Ensure a sufficient amount of DNA is used as indicated in Step 3.
	Little or no 8-OHdG in the sample.	N/A
Uneven fluorescent development	Delayed fluorescence development in the wells.	Ensure fluorescence development solution is added sequentially and is consistent with the order you added the other reagents (e.g., from well A to well G or from well 1 to well 12). If using 12 or more wells, a multi-channel pipette should be used so that the addition of fluorescence development solution can be completed as quickly as possible (<20 seconds).

## RELATED PRODUCTS

### DNA Sample Preparation

P-1003	FitAmp™ General Tissue Section DNA Isolation Kit
P-1004	FitAmp™ Plasma/Serum DNA Isolation Kit
P-1006	DNA Concentrator Kit
P-1009	FitAmp™ Paraffin Tissue Section DNA Isolation Kit
P-1017	FitAmp™ Urine DNA Isolation Kit
P-1018	FitAmp™ Blood and Cultured Cell DNA Extraction Kit

### Methylated and Hydroxymethylated DNA Quantification

P-1034	MethylFlash™ Methylated DNA Quantification Kit (Colorimetric)
P-1035	MethylFlash™ Methylated DNA Quantification Kit (Fluorometric)
P-1036	MethylFlash™ Hydroxymethylated DNA Quantification Kit (Colorimetric)
P-1037	MethylFlash™ Hydroxymethylated DNA Quantification Kit (Fluorometric)

### Hydroxymethylated DNA Immunoprecipitation

P-1038	MethylFlash™ Hydroxymethylated DNA Immunoprecipitation (hmeDIP) Kit
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### DNA Damage and Repair

OP-0001	EpiQuik™ Superoxide Dismutase Activity/Inhibition Assay Kit (Colorimetric)
OP-0004	CytoX-Red Cell Proliferation/Cytotoxicity Assay Kit
OP-0005	CytoX-Violet Cell Proliferation/Cytotoxicity Assay Kit
P-6003	EpiQuik™ 8-OHdG DNA Damage Quantification Direct Kit (Colorimetric)
P-6001	EpiQuik™ In Situ DNA Damage Assay Kit

### DNA Damage and Repair Antibodies

(See [http://www.epigentek.com/catalog/dna-damage-repair-c-35\\_70.html](http://www.epigentek.com/catalog/dna-damage-repair-c-35_70.html))