EpiQuik™ Global Acetyl Histone H3-K36 Quantification Kit (Fluorometric)

Base Catalog # P-4019

PLEASE READ THIS ENTIRE USER GUIDE BEFORE USE

The *EpiQuik™* Global Acetyl Histone H3-K36 Quantification Kit (Fluorometric) is suitable for specifically measuring global histone H3-K36 acetylation using a variety of mammalian cells (human, mouse, etc.) including fresh and frozen tissues, cultured adherent and suspension cells.

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KIT CONTENTS

Components	48 assays P-4019-48	96 assays P-4019-96
F1 (10X Wash Buffer)	10 ml	20 ml
F2 (Antibody Buffer)	6 ml	12 ml
F3 (Detection Antibody, 1 mg/ml)*	5 µl	10 µl
F4 (Fluoro Developer)*	12 µl	24 µl
F5 (Fluoro Enhancer)*	12 µl	24 µl
F6 (Fluoro Dilution)	4 ml	8 ml
Standard Control (100 µg/ml)*	10 µl	20 µl
8-Well Sample Strips (with Frame)	4	9
8-Well Standard Control Strips	2	3
User Guide	1	1

** For maximum recovery of the products, centrifuge the original vial prior to opening the cap.*

SHIPPING & STORAGE

Upon receipt, store F3, F4 and Standard control at –20°C away from light. Store all other components at 4°C away from light. The components of the kit should be stable for 6 months when stored properly.

Note: Check if buffers F1 and F2 contain salt precipitates before using. If so, warm (at room temperature or 37°C) and shake the buffers until the salts are re-dissolved.

MATERIALS REQUIRED BUT NOT SUPPLIED

- ☐ Orbital shaker
- ☐ Pipettes and pipette tips
- ☐ Reagent reservoir
- ☐ Fluorescence microplate reader
- ☐ 15 ml conical tube
- ☐ 1.5 ml microcentrifuge tubes

GENERAL PRODUCT INFORMATION

Usage Limitation: The *EpiQuik*™ Global Acetyl Histone H3-K36 Quantification Kit (Fluorometric) is for research use only and is not intended for diagnostic or therapeutic application.

Safety: Suitable lab coat, disposable gloves, and eye protection are required when working with the kit.

Quality Control: Epigentek guarantees the performance of all products in the manner described in our product instructions.

Product Updates: Epigentek reserves the right to change or modify any product to enhance its performance and design.

Intellectual Property: The *EpiQuik*™ Global Acetyl Histone H3-K36 Quantification Kit (Fluoriometric) and methods of use contain proprietary technologies by Epigentek. *EpiQuik*™ is a trademark of Epigentek Group Inc.

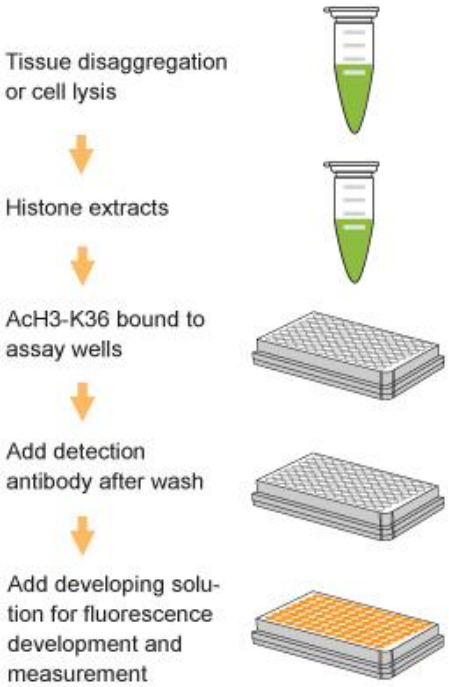
A BRIEF OVERVIEW

Acetylation of histones including histone H3 has been involved in the regulation of chromatin structure and recruitment of transcription factors to the gene promoters. Histone acetyltransfeases (HAT) and histone deacetylases (HDACs) play a critical role in control of histone H3 acetylation at multiple sites. Acetylation of histone H3 at lysine 36 (H3-K36) is a conserved modification which functions in transcription. Acetylation of histone H3-K36 is tightly involved in the cell cycle reguation, cell proliferation and apoptosis. An imbalance in the equilibrium of histone H3 acetylation including K36 acetylation has been associated with cancer progression. Histone H3-K36 acetylation may be increased by inhibition of HDACs and decreased by HAT inhibition. Thus quantitative detection of global acetyl histone H3-K36 would provide useful information for better understanding epigenetic regulation of gene activation and for developing HAT or HDAC-targeted drugs. The *EpiQuik*™ Global Acetyl Histone H3-K36 Quantification Kit (Fluorometric) provides a tool for measuring global acetylation of histone H3-K36. The kit has the following features:

- Quick and efficient procedure, which can be finished within 2.5 hours.
- Innovative fluorometric assay with no need for radioactivity, electrophoresis, and chromatography.
- Specifically capturing acetyl H3-K36 with the detection limit as low as 0.4 ng/well and detection range from 5 ng-2 μg/well of histone extracts.
- The control is conveniently included for quantification of the amount of acetyl H3-K36.
- Strip microplate format makes the assay flexible: manual or high throughput.
- Simple, reliable, and consistent assay conditions.

PRINCIPLE & PROCEDURE

The *EpiQuik*™ Global Acetyl Histone H3-K23 Quantification Kit (Fluorometric) is designed for measuring global histone H3-K36 acetylation. In an assay with this kit, the acetyl histone H3 at lysine 36 is captured to the strip wells coated with anti-acetyl H3-K36 antibody. The captured acetyl histone H3-K36 can be then detected with a labeled detection antibody followed by fluorescent development reagent. The ratio of acetyl H3-K36 is proportional to the intensity of fluorescence. The absolute amount of acetyl H3-K36 can be quantified by comparing to the standard control.



Schematic Procedure for Using the *EpiQuik*™ Global Acetyl Histone H3-K36 Quantification Kit (Fluorometric)

PROTOCOL

1. **a)** Prepare histone extracts from cells/tissues treated or untreated by using your own successful method (acid extraction or high salt extraction).
   b) For your convenience and the best results, Epigentek offers the Fast Total Histone Extraction kit (Cat # OP-0006) optimized for use in the *EpiQuik*™ modified histone quantification series.
   c) Preparation of histone extracts can also be performed using the attached procedure (See Appendix). Histone extracts can be used immediately or stored at –80°C for future use.
2. Determine the number of the strip wells required. Leave these strips in the plate frame (remaining unused strips can be put back in the bag. Seal the bag tightly and store at 4°C). Dilute **F1** with distilled water (pH 7.2-7.5) at the 1:9 ratios (1 ml of **F1** + 9 ml of distilled water)
3. Add 50 µl of **F2** into each well. For the sample, add 50-200 ng of the histone extract into the sample wells. For standard curve, dilute **standard control** with **F2** to 1 – 100 ng/µl for 5-7 points (ie: 1.5, 3, 6, 12, 25, 50, and 100 ng/µl). Add 1 µl of **standard control** at the different concentrations into the standard well**.** For the blank, add no nuclear extracts or no standard control protein. Mix and cover the strip wells with Parafilm M and incubate at room temperature for 1-2 hours.
4. Aspirate and wash the wells with 150 µl of **diluted F1** 3 times.
5. Dilute **F3** (at 1:1000 ratio) to 1 µg/ml with **F2**. Add 50 µl of diluted **F3** to each well and incubate at room temperature for 60 min on an orbital shaker (100 rpm).
6. Aspirate and wash the wells with 150 µl of **diluted F1** 6 times.
7. Prepare **fluoro-development solution** by adding 1 µl of **F4** and 1 µl of **F5** into each 400 µl of **F6**. Add 50 µl of **fluoro development soultion** into the wells and incubate at room temperature for 1-5

min away from light. The color in the standard wells containing the higher concentrations may turn slightly pink during this period. Measure and read fluorescence on fluorescence microplate reader at $530_{EX}/590_{EM}$ nm.

Note: If the strip well frame does not fit the fluorescence reader, transfer the solution to the standard 96-well microplate and read fluorescence at $530_{EX}/590_{EM}$ nm.

8. Calculate % histone H3-K36 acetylation:

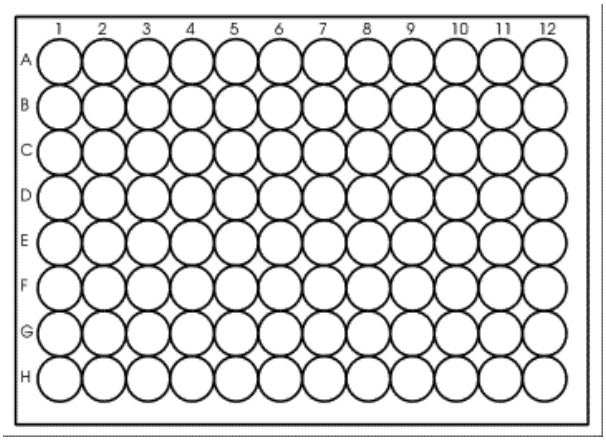
$$\text{Acetylation \%} = \frac{\text{RFU (treated (tested) sample – blank)}}{\text{RFU (untreated (control) sample – blank)}} \times 100\%$$

For the amount quantification, plot RFU versus amount of **standard control** and determine the slope as delta RFU/ng.

Calculate the amount of acetyl H3-K36 using the following formula:

$$\text{Amount (ng/mg protein)} = \frac{\text{RFU (sample – blank)}}{\text{slope}} \times 1000$$

PLATE CONFIGURATION



- **Strip 1-3 (for 96 assays) or strip 1-2 (for 48 assays)**: standard wells (**green trimmed**); The standard curve can be generated with 5-8 concentration points (includes blank).
- Example amount of standard control /well: **A1**: 100 ng; **B1**: 50 ng; **C1**: 25 ng; **D1**: 12 ng; **E1**: 6 ng; **F1**: 3 ng; **G1** 1.5 ng; **H1**: 0 ng
- **Strip 4-12 (for 96 assays) or strip 3-6 (for 48 assays):** sample wells (**No label**)
- Each sample or standard point can be assayed in the duplicates or triplicates.

Appendix

Histone Extraction Protocol

1. *For tissues (treated and untreated),* weigh the sample and cut the sample into small piece (1-2 mm^3) with a scalpel or scissors. Transfer tissue pieces to a Dounce homogenizer, add TEB buffer (PBS containing 0.5% Triton X 100, 2 mM PMSF and 0.02% NaN_3) at 200 mg/ml, and disaggregate tissue pieces by 50-60 strokes. Transfer homogenized mixture to a 15 ml conical tube and centrifuge at 3,000 rpm for 5 min at 4°C. If total mixture volume is less than 2 ml, transfer mixture to a 2 ml vial and centrifuge at 10,000 rpm for 1 min at 4°C. Remove supernatant.

   For cells (treated and untreated), harvest cells and pellet the cells by centrifugation at 1000 rpm for 5 min at 4°C. Resuspend cells in TEB buffer at 10^7 cells/ml and lyse cells on ice for 10 min with gentle stirring. Centrifuge at 3000 rpm for 5 min at 4°C. If total volume is less than 2 ml, transfer cell lysates to a 2 ml vial and centrifuge at 10000 rpm for 1 min at 4°C. Remove supernatant.

2. Resuspend cell/tissue pellet in 3 volumes (approx. 200 μl/10^7 cells or 200 mg tissues) of extraction buffer (0.5N HCl + 10% glycerol) and incubate on ice for 30 min.

3. Centrifuge at 12,000 rpm for 5 min at 4°C and remove the supernatant fraction to new vial.

4. Add 8 volumes (approx. 0.6 ml/ 10^7 cells or 200 mg tissues) of acetone and leave at –20°C overnight.

5. Centrifuge at 12,000 rpm for 5 min and air-dry the pellet. Dissolve the pellet in distilled water (30-50 μl/10^7 cells or 200 mg tissues).

6. Quantify the protein concentration. Aliquot the extract and store the extract at –20°C or –80°C.

TROUBLESHOOTING

No Signal for Both the Standard Control and the Samples

Reagents are added incorrectly.	Check if reagents are added in order and if any steps of the procedure may have been omitted by mistake.
Incubation time and temperature are incorrect.	Ensure the incubation time and temperature described in the protocol are followed correctly.

No Signal or Very Weak Signal for Only the Standard Control

The amount of standard control is not added into "standard control wells" or is added insufficiently.	Ensure sufficient amount of control is properly added to the standard control wells.

No Signal for Only the Sample

The protein sample is not properly extracted.	Ensure the procedure and reagents are correct for the nuclear protein extraction.
The protein amount is added into well insufficiently.	Ensure extract contains sufficient amount of protein.
Protein extracts are incorrectly stored.	Ensure the nuclear extracts are stored at –20°C or –80°C.

High Background Present for the Blank

The well is not washed enough.	Check if wash at each step is performed according to the protocol.
Contaminated by the standard control.	Ensure the well is not contaminated from adding the control protein or from using control protein contaminated tips.
Overdevelopment.	Decrease development time in Step 7

RELATED PRODUCTS

P-4010	*EpiQuik*™ Global Acetyl Histone H3-K9 Quantification Kit (Colorimetric)
P-4011	*EpiQuik*™ Global Acetyl Histone H3-K9 Quantification Kit (Fluorometric)
P-4012	*EpiQuik*™ Global Acetyl Histone H3-K14 Quantification Kit (Colorimetric)
P-4013	*EpiQuik*™ Global Acetyl Histone H3-K14 Quantification Kit (Fluorometric)
P-4014	*EpiQuik*™ Global Acetyl Histone H3-K18 Quantification Kit (Colorimetric)
P-4015	*EpiQuik*™ Global Acetyl Histone H3-K18 Quantification Kit (Fluorometric)
P-4016	*EpiQuik*™ Global Acetyl Histone H3-K23 Quantification Kit (Colorimetric)
P-4017	*EpiQuik*™ Global Acetyl Histone H3-K23 Quantification Kit (Fluorometric)